

RIDL; What Is It? How Does It Work? Does It Work? And What's In The Future...?

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Background: Oxitec Limited

- Developing environment-friendly technology to control insect pests since 2002
 - Agriculture
 - Human Health
- Building on innovative research from Oxford University
- International recognition with a \$ 5 million grant from the Gates Foundation Grand Challenges for Global Health Initiative



Male medfly



Aedes albopictus

What is RIDL?

RIDL® Release of Insects with Dominant Lethality

Innovative and environment-friendly insect control technology

□ Genetic enhancement of the Sterile Insect Technique (SIT)

■ Targeting insect pests that are damaging to agriculture and human health worldwide

Pest control using GM insects

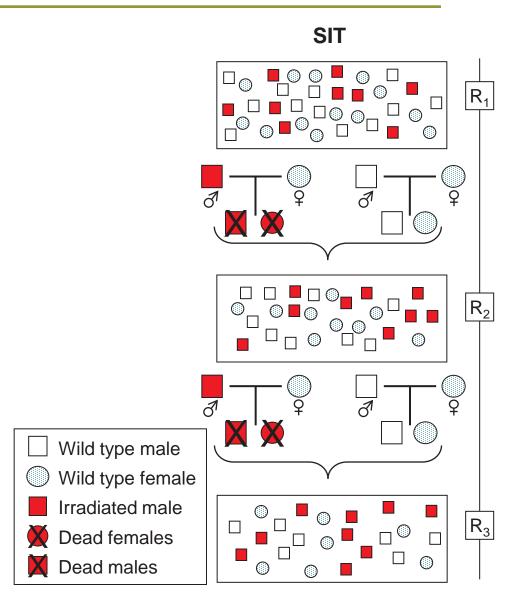
Population suppression
 Goal: reduce numerical size of pest population

"Population replacement" or "Refractory insect strategy"

Goal: change pest population to less harmful form

Sterile Insect Technique (SIT)

- Rear millions of insects
- Sterilize (usually with irradiation)
- Release over wide area
- Native females that mate with sterile males produce non viable eggs
- target pest population declines



Success of SIT

- SIT has been successfully used to eradicate/control;
 - New World Screwworm.
 - Mediterranean fruit fly.
 - Melon fly from Okinawa islands of Japan.
 - Queensland fruit fly in Australia.
 - Mexican fruit fly in Northern Mexico.
 - West Indian fruit fly in Northern Mexico.
 - Codling Moth in Canada.
 - Pink Bollworm in California.

Background: SIT and mosquitoes

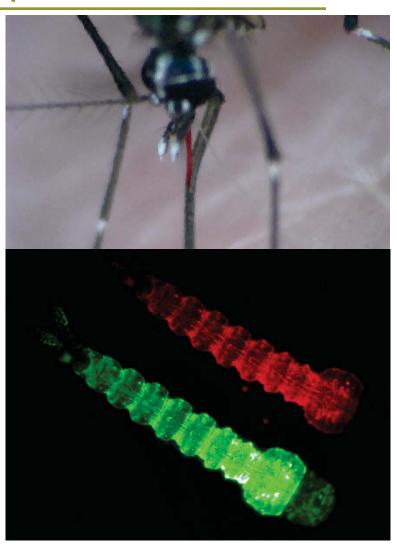
□ >20 trials in 1970's - 1980's

[Reviewed by Mark Benedict and Alan Robinson (2003) *Trends Parasitol.* 19:349]

- Variable success
- Some problems revealed:
 - Immigration of pre-mated females
 - Poor competitiveness of irradiated/chemosterilised males
 - Breakdown and semi-sterility of strains using translocations or other chromosomal rearangments.

A Genetic Solution for mosquito control

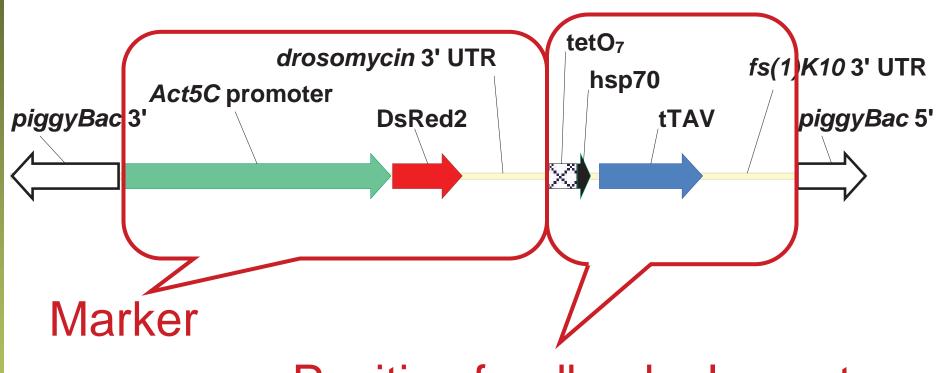
- Based on Sterile Insect Technique
- Uses modern molecular biology techniques to insert a lethal gene into insects
- Sterile transgenic insects can be used to control a range of pests.



"birth control for mosquitoes"

How does it work?

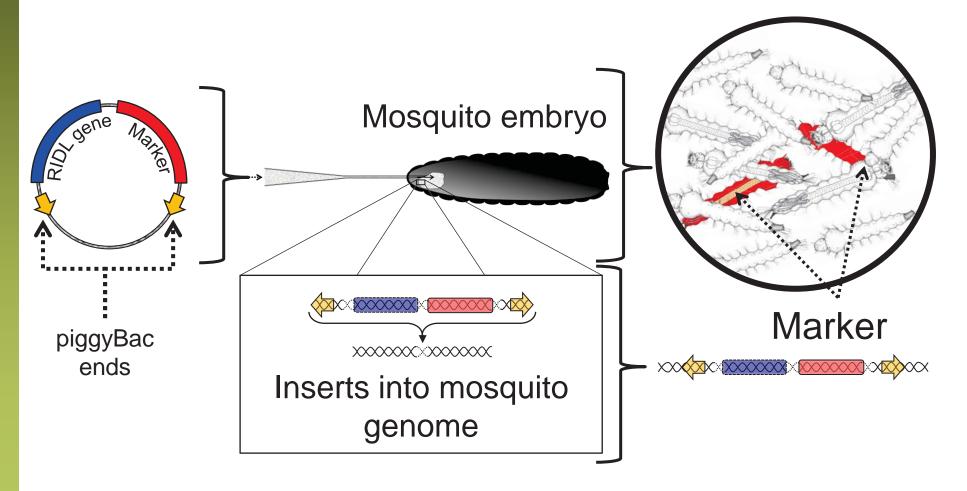
OX513 - structure



Positive feedback element

Microinjection

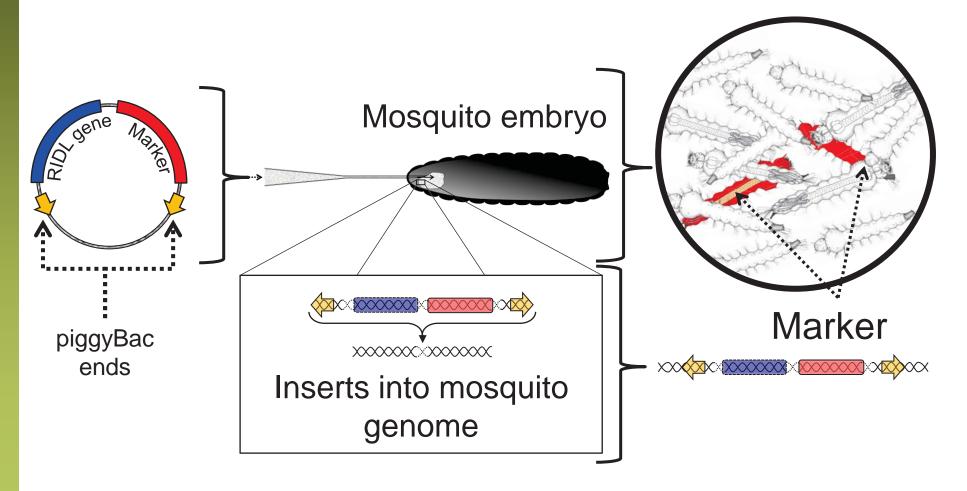
■ Produce RIDL line





Microinjection

■ Produce RIDL line

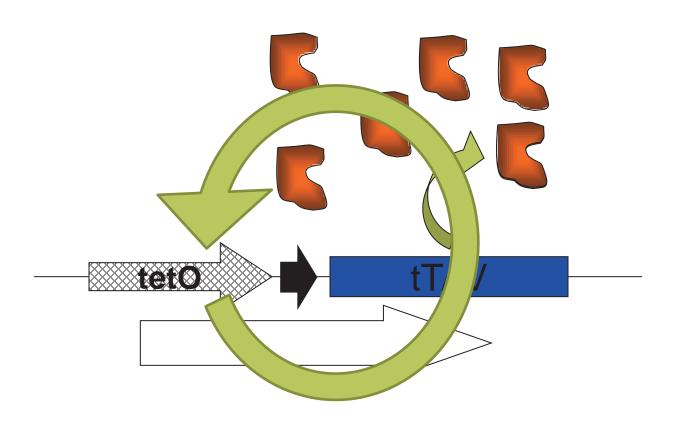


OX513A – Fluorescent marker



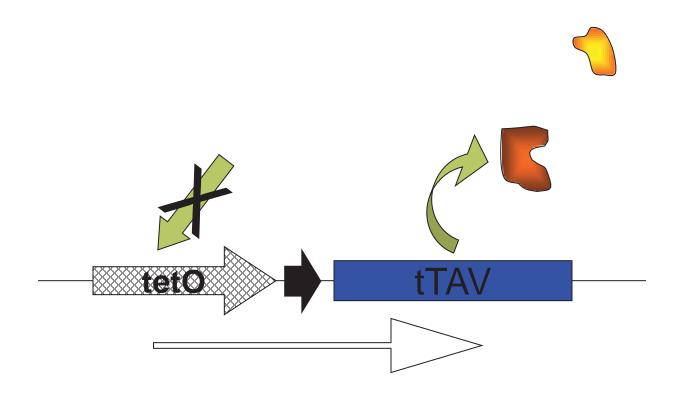
RIDL gene function

□ Bi-sex RIDL is based on positive feedback system;



RIDL gene control

□ Bi-sex RIDL is based on positive feedback system;



RIDL; summary of mode of action

- Accumulation of tTA in the cells causes insect death (transcriptional squelching): tTA is a lethal gene
- The accumulation of tTA is repressed by the presence of tetracycline: the lethality is conditional
- One copy of the tTA gene in the genome is enough to kill the insect: the lethal gene is dominant

Current tools for vector control

approach	required citizen compliance	advantages	disadvantages	best fit
Fogging/space spraying	high	quick kill	contact requiredhuman exposureineffective	rapid responsesmall location
Larvicides	high	duration (weeks)	impossible to treat all breeding sitesdrinking water contamination	programmatic activitypre season
Lethal ovitraps	high	home treatment	costreplenishmentefficacy issues	• unproven
Aerosols, mats, coils, treated fabrics	high	in door use	• individual	• top up

High public compliance reliance + transient effect + 'contact cost' + insecticide resistance = global failure to arrest spread of dengue

Traditional approaches

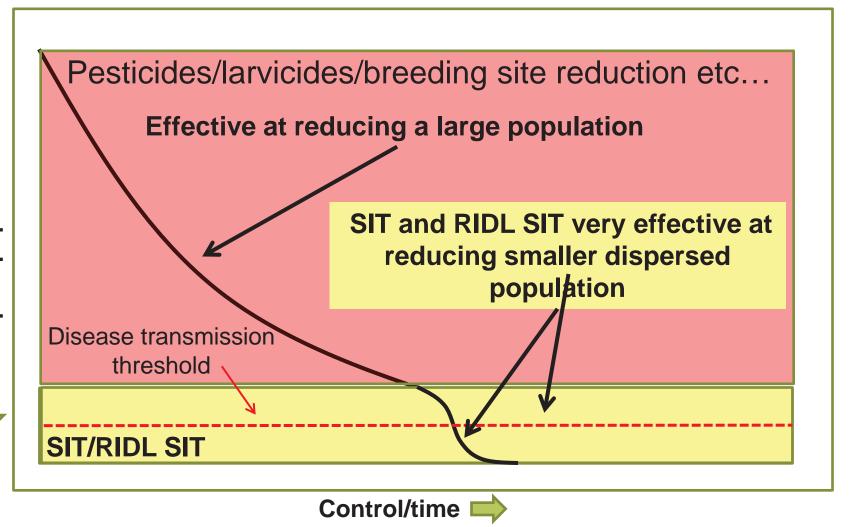


Fogging – areas Missed by vehicle

traditional approaches



Larviciding or placing lethal ovitraps by hand



Trial process

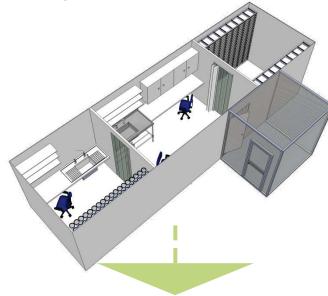
Filter Colony (Oxitec UK)

- Hybrid strain
- High Quality control
- Fitness
- Genetic diversity
- Longevity

Eggs sent to field site



Rearing cabin / factory



Monitoring



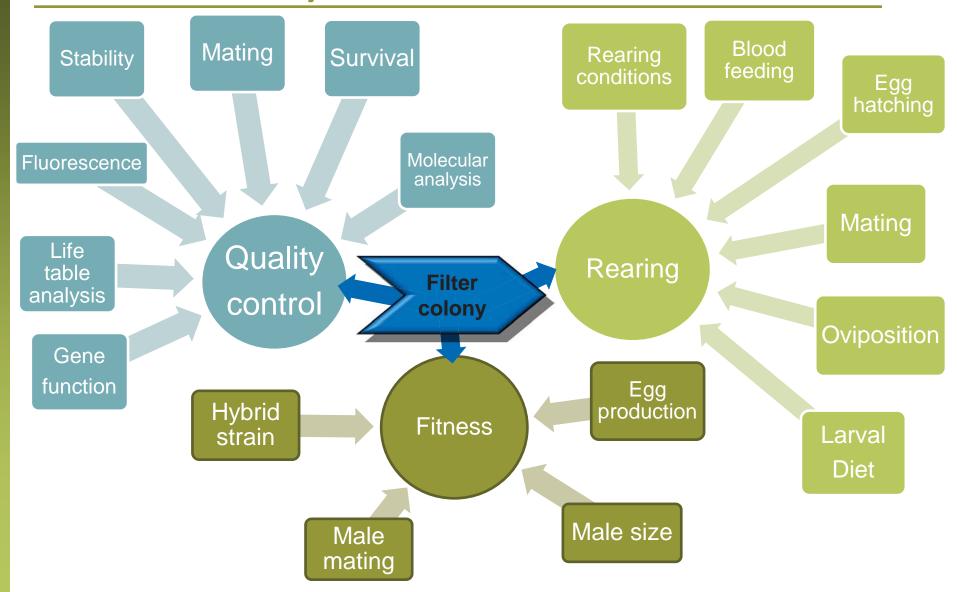
Release by vehicle /hand



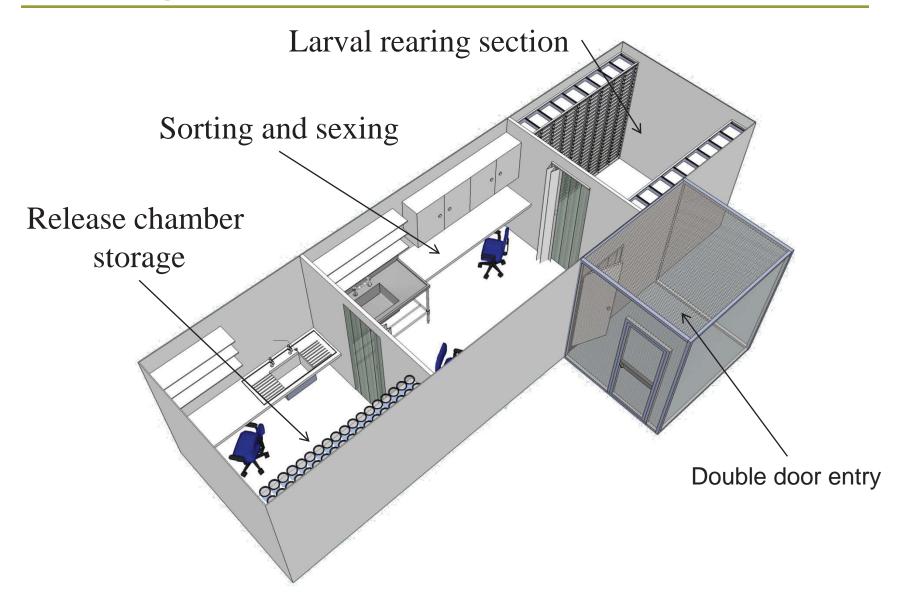


Sex-sort pupae
Males for release

Filter colony UK - overview

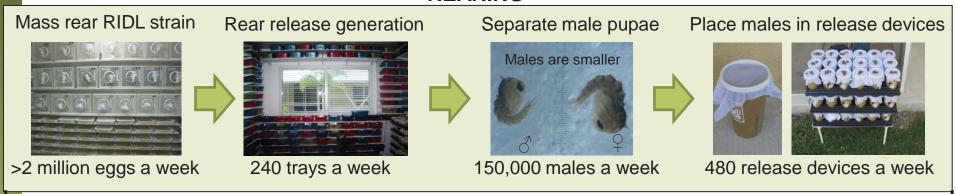


Rearing facility



2010 suppression trial; methods

REARING



(3 people)

DISTRIBUTION



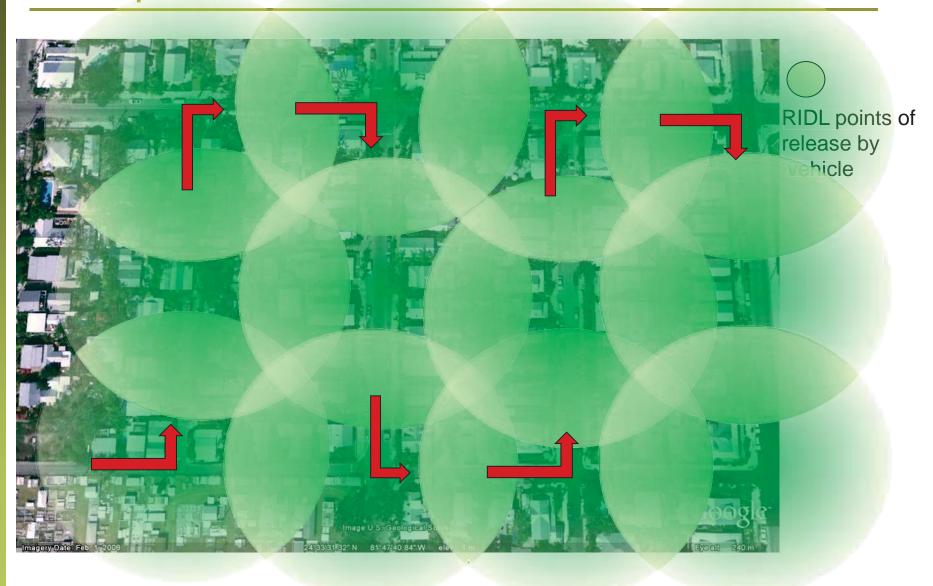


MONITORING Adults



(2 people)

Trial process: Distribution

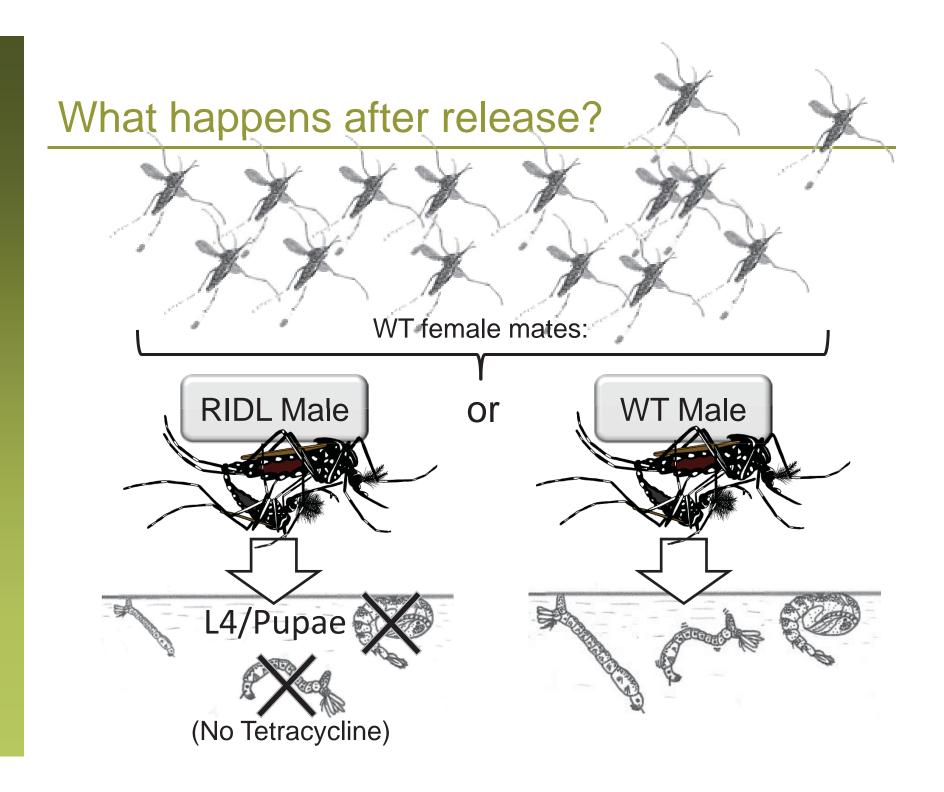


Trial process: Monitoring

Monitoring

Monitor using ovitraps (200) and BG traps (40)





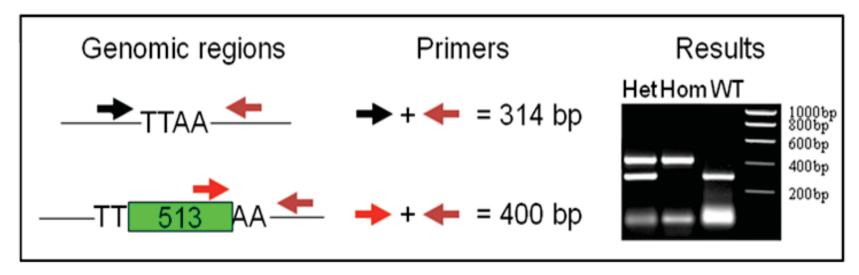
Does it work?

Strain testing

- Several strains tested for the following;
 - Molecular characterisation
 - Phenotype
 - Mating competitiveness
 - Life table parameters
 - Gene stability
 - Insecticide resistance
 - Effects on predators
- Field testing
 - Cayman
 - Brazil

OX513A molecular characterisation

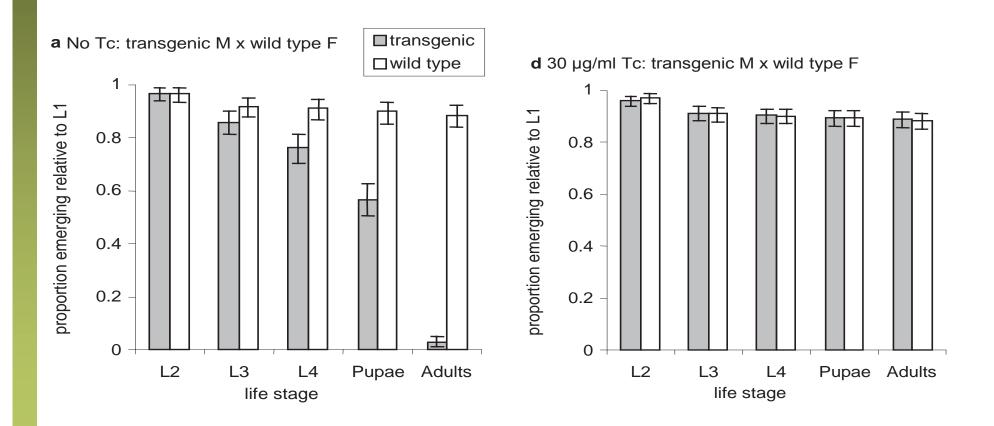
- OX513A was created in 2002, injected into a Rockefeller background
- Single integration event (Phenotypic and molecular analysis)



■ Insertion has been sequenced

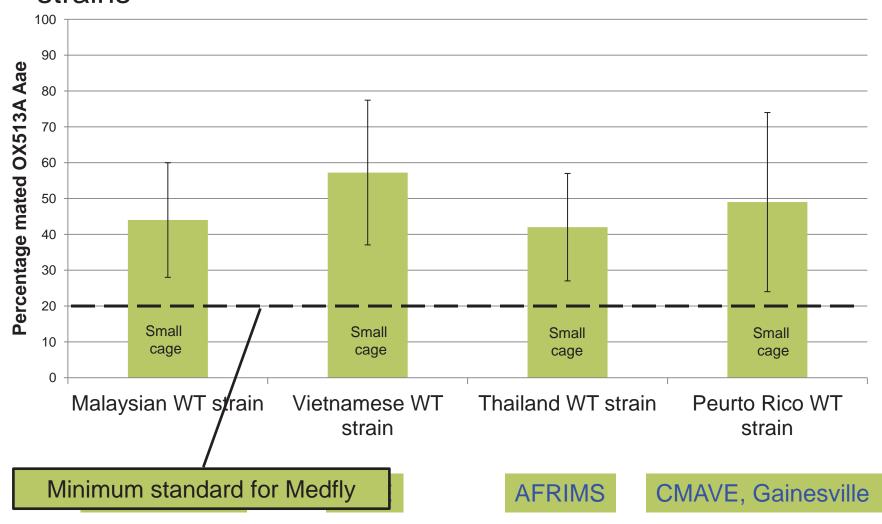
OX513A Aae phenotypic analysis

Penetrance of RIDL in OX513A Aae



Contained mating trials results

Percentage mating of OX513A Aae and different WT strains

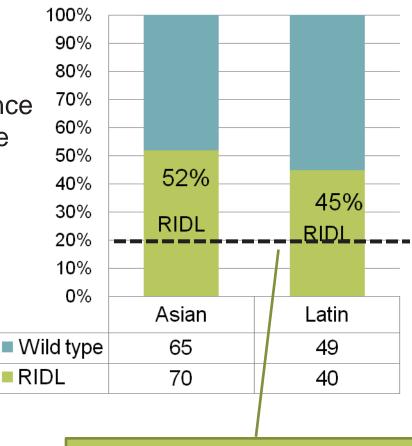


Lab & Field House (Contained) Testing Results

- Laboratory; No significant difference between OX513A and control wild type strain in all life history parameters
- Field House; No statistical difference in mate selection between wild type and RIDL of either background



Number of females that mated with RIDL males



Minimum standard for Medfly

OX513A Life Table comparison to WT

Parameter	Strain	Average	Mean Rank	p-Value
Eggs laid per female	RIDL	74.86 ± 16.94	16.03	0.740
	wild type	63.07 ± 6.855	14.97	
Larvae hatched per egg batch.	RIDL	49.86 ± 8.306	17.70	0.171
	wild type	31.60 ± 5.237	13.30	
Sterile eggs per egg	RIDL	25.00 ± 3.349	14.33	0.468
batch	wild type	31.47 ± 5.57	16.67	
Days spent in L4	RIDL	2.59 ± 0.06	142.84	0.851
	wild type	2.57 ± 0.58	141.20	
Days spent at pupal stage	RIDL	2.29 ± 0.042	133.52	0.141
	wild type	2.36 ± 0.40	144.99	
Days from hatching	RIDL	7.85 ± 0.071	130.19	0.200
to adult	wild type	7.49 ± 0.055	141.28	

OX513A Gene stability

- Stable for over 60 generations as a homozygous line
 - Molecular and marker expression analysis
- Re-exposure to transposase does not cause movement at detectable levels
- Penetrance of phenotype unchanged

OX513A strain background

Insecticide resistance of OX513A Aae Latin and Latin strain background has been tested

Insecticide	Latin background strain	OX513A Aae Latin strain	Resistant
Temephos	95%	100%	NO
Deltamethrin	99%	100%	NO
Permethrin	100%	100%	NO
Bendiocarb	73%	47%	YES
Malathion	100%	100%	NO

- Bendiocarb no longer used in US (too toxic)
- No kdr resistant alleles were found;
 - □ (Tetraplex (1534), Hola (Val1016lle), F1534C and V1016G tested)

OX513A Aae effects on predators

- Toxorhynchites; predatory species, larvae feed exclusively on other mosquito larvae
- □ Fed two different species of Toxorhynchites on OX513A Aae and WT larvae
- Found no significant difference in any of the following;
 - Larval development
 - Lifespan
 - Size
 - Fecundity
 - Survival of any life stage
- Study conducted by IMR, Malaysia

2010 Cayman Suppression Trial

Aims of the trial

- Increase in Male: Female ratio
 - Aimed for 10:1, based on SIT and models
- Mating competitiveness of RIDL males
 - Fluorescent genetic marker in larvae hatched from eggs collected in Ovitraps
 - **>50%**
- Suppression of target mosquito population
 - Ovitrap index
 - Significant and sustained difference between treated and control

Field site (East End)

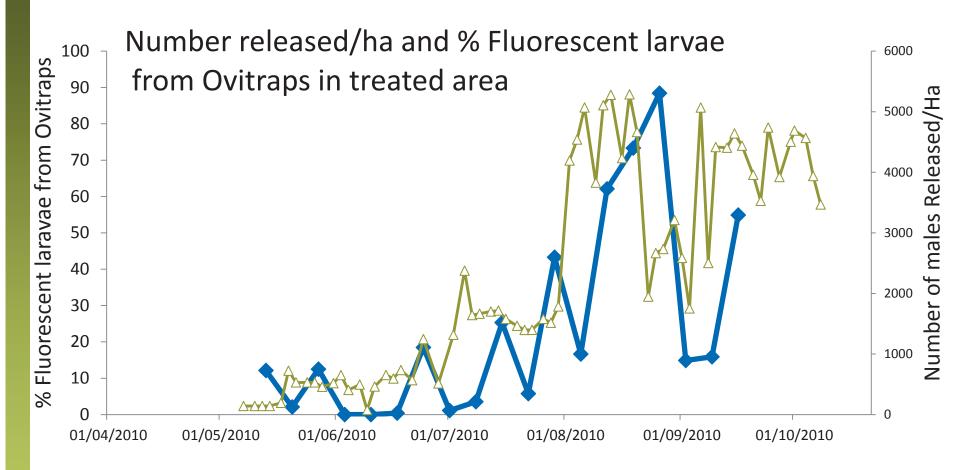


Each Area approx 16 Ha (40 acres)

No conventional control for Aedes aegypti

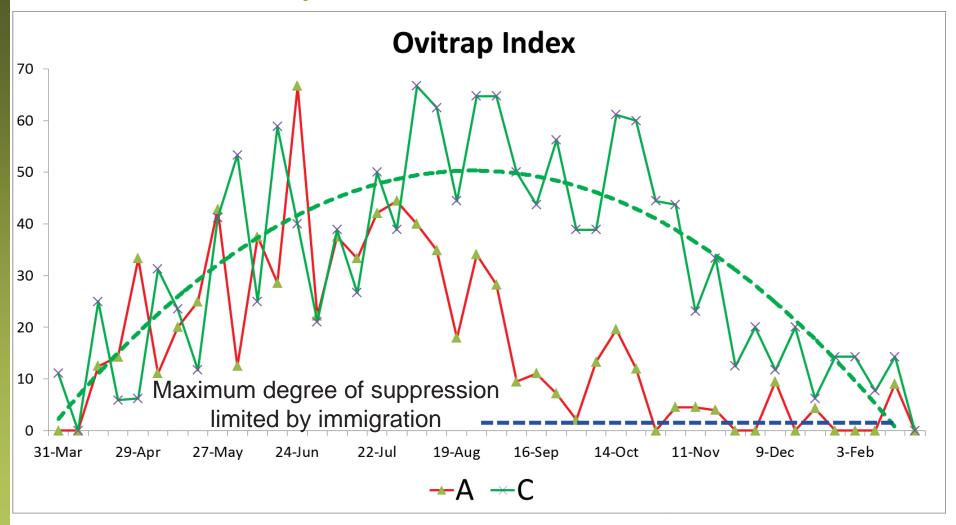
Release period May-October with pre- and post-release monitoring

Results - Fluorescent score



- Fluorescence increases in line with, but lagging, male releases
- Later dates had too few eggs to calculate ratio

Results: Ovitrap index



Clear suppression from early August

Cayman field trial summary

- Trial was successful
 - All endpoints successfully met
 - Sterile:wild male ratio in traps >10:1
 - □ Fluorescence ratio in progeny >50%
 - Population suppression
 - □ Sterile:wild male ratio 3-4:1 sufficient
- Sustained release of RIDL OX513A males can suppress a field population of Aedes aegypti mosquitoes
- □ GE mosquitoes can perform successfully in the field

2012; results from Brazil

■ Brazil is a collaboration with;



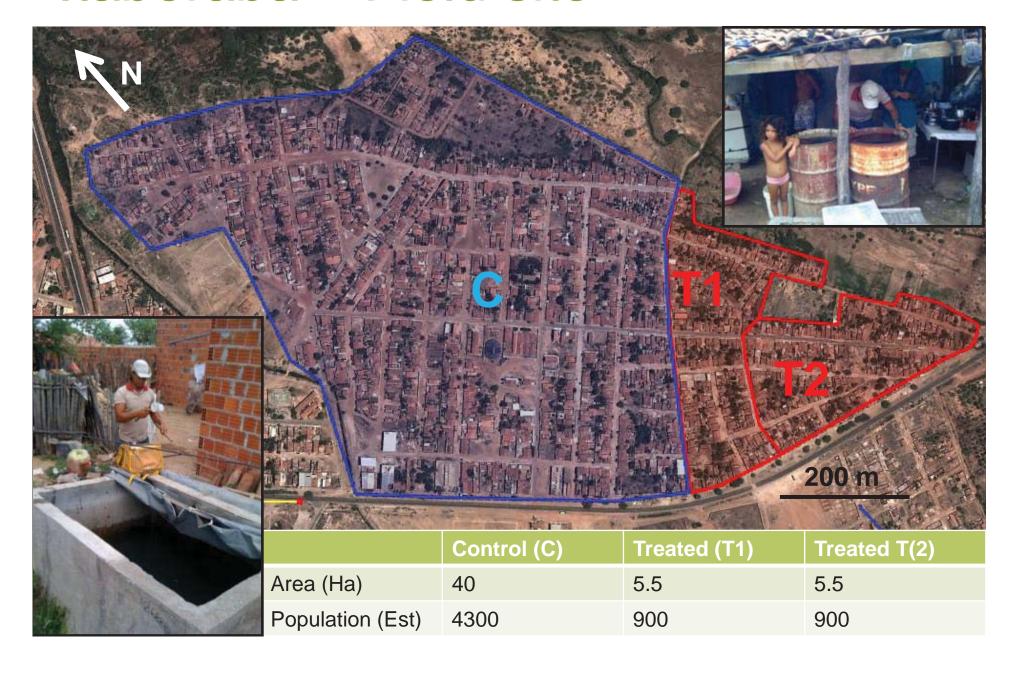




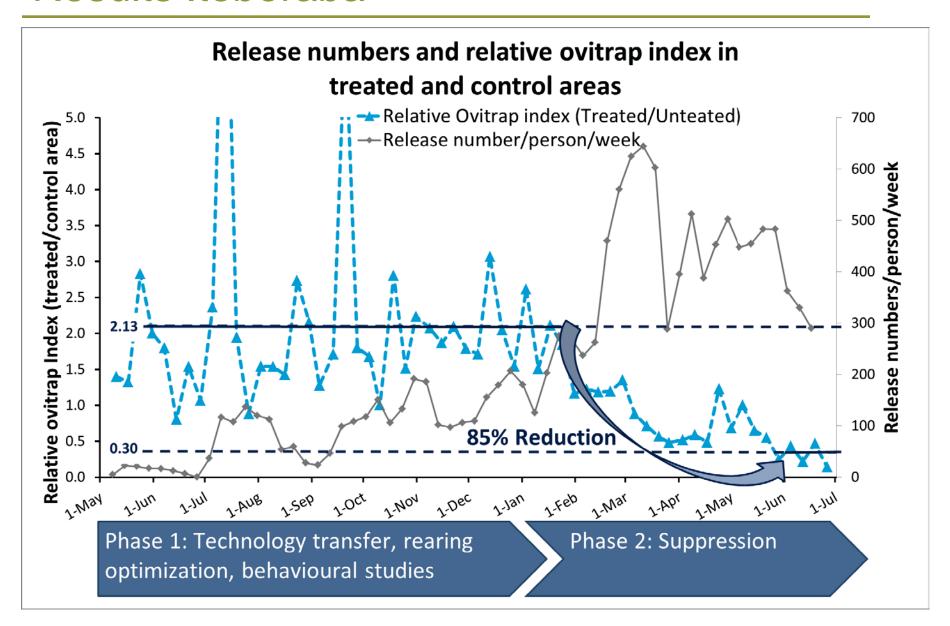
- Working in two trial sites for the past year
 - Iteberaba
 - Mandacaru



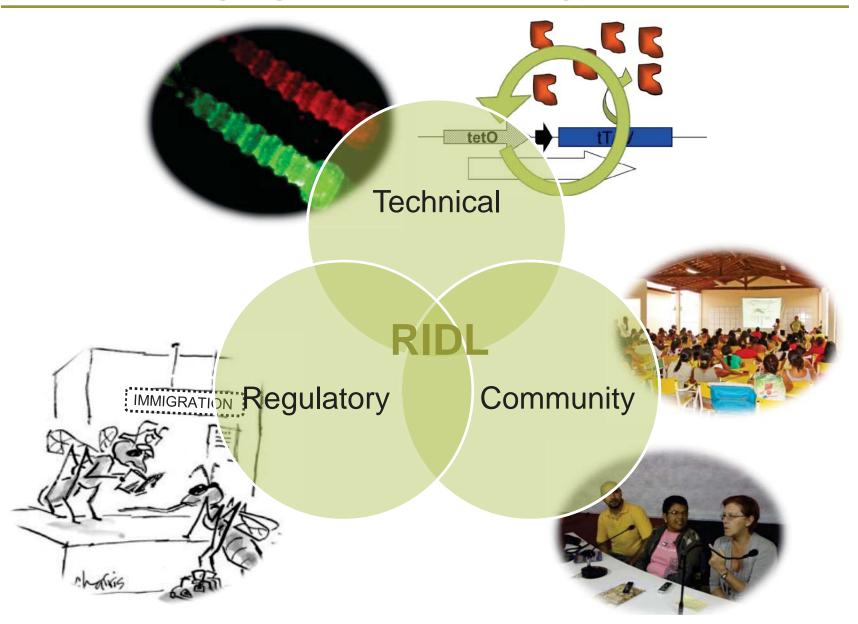
Itaberaba – Field site



Results Iteberaba



RIDL® bringing new technology to the field



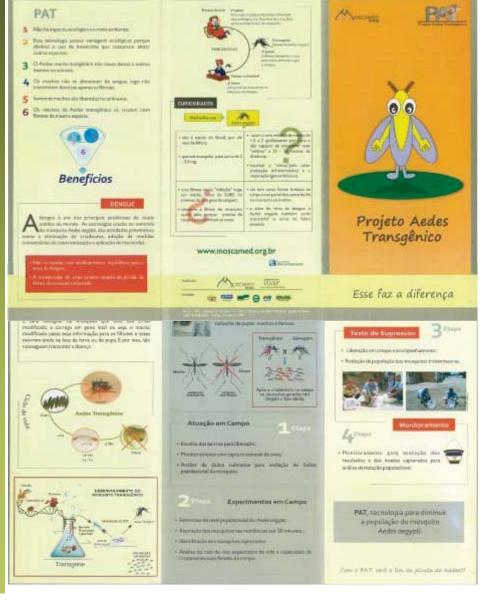
Regulatory process

- Regulatory framework is evolving, with trials approved and conducted in Malaysia, Brazil, Cayman.
- Even authorities with extensive experience in biotechnology regulation have to adapt frameworks for GE insect regulation
 - US process for a trial started in 2008
 - USDA-VS issued letter of no jurisdiction after 18 months deliberation
 - FDA and EPA still discussing who should regulate GE mosquitoes.

Community Engagement

- Community engagement plans are in place for each trial
- Main avenues of community engagement;
 - Radio interviews and local media
 - Public information events; for example public outreach to schools, guilds and hospitals, leaflets
 - Consultation with senators; through the regulatory process and communicating with local senators.
 - Web site information (local mosquito control district)

Community engagement





- * Female mosquitoes mate only once
- If a wild female mates with a sterile male then she will not be able to produce offspring
- *We are releasing sterile male mosquitoes as part of a new research project
- # if many females mate with our sterile males they will produce no offspring and there will be a reduction in the Dengue Mosquito population
- * Male mosquitoes don't bite only the females blood feed in order to use the protein to make their eggs



For more information Contact:

The Morquite Research & Control Unit
(345) 919-2457

Female Mosquitoes mate just once in their lifetime



Usually this means hundreds or thousands of offspring

...but if the Wild Female mates a Sterile Male



Then she has no children!

MOSQUITO



RESEARCH & CONTROL UNIT

We are currently testing a new technology to reduce numbers of the



MRCU personnel are working in East End on a new project

these traps to monitor our success with this new project



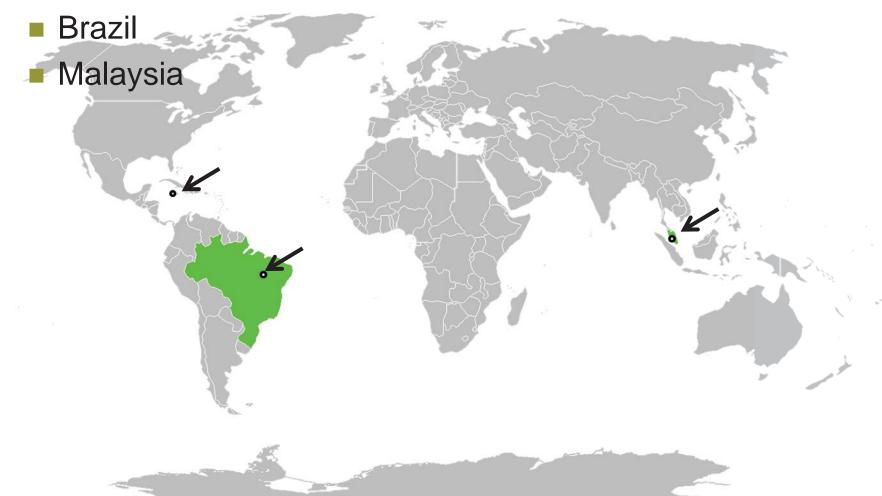
If you see this equipment please do not disturb it



What's in the future?

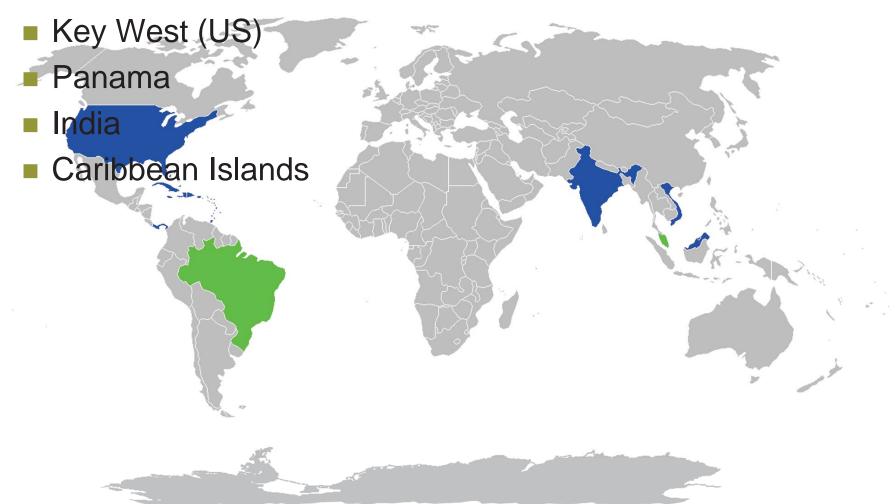
On-going control programmes

- Open field releases conducted so far;
 - Cayman Islands

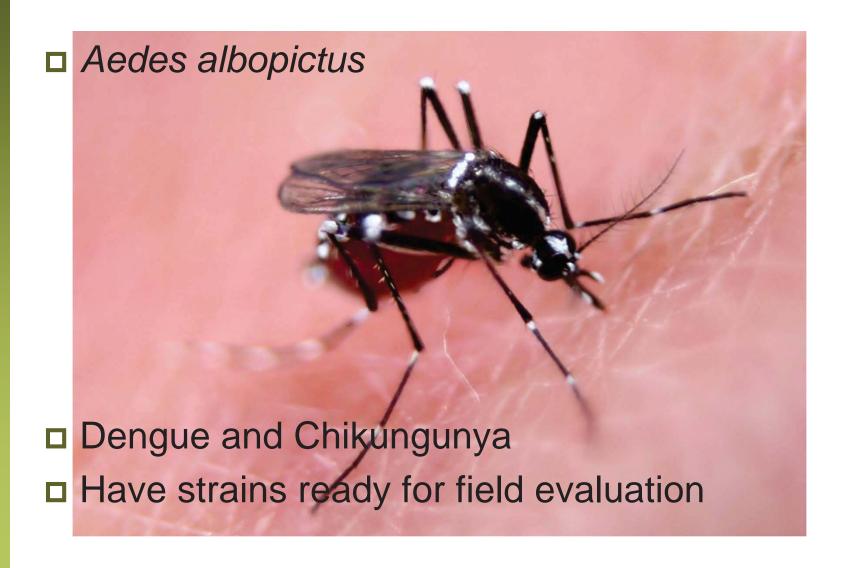


On-going control programmes

Currently seeking regulatory approval to use RIDL;



New species (mosquitoes)



New species (mosquitoes)

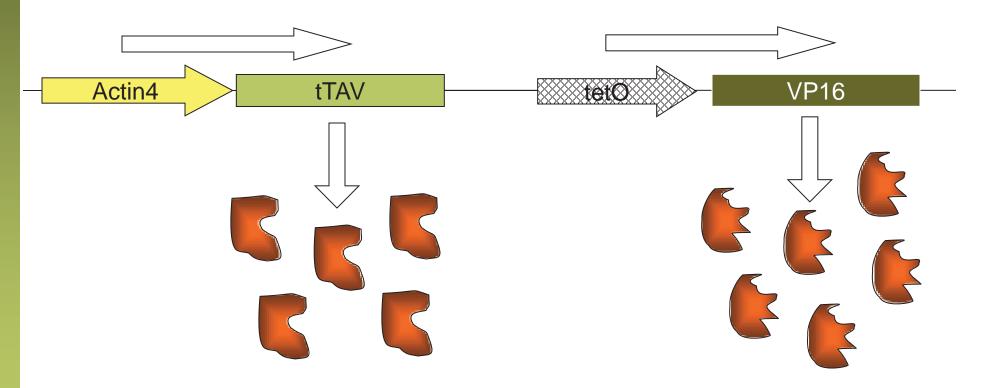
Culex sp.

□ SBIR grant to transform Culex quinquefasciatus (C. pipiens and C. tarsalis)

Ultimate aim to develop RIDL strains for the control of WNV

Bi-sex RIDL technology

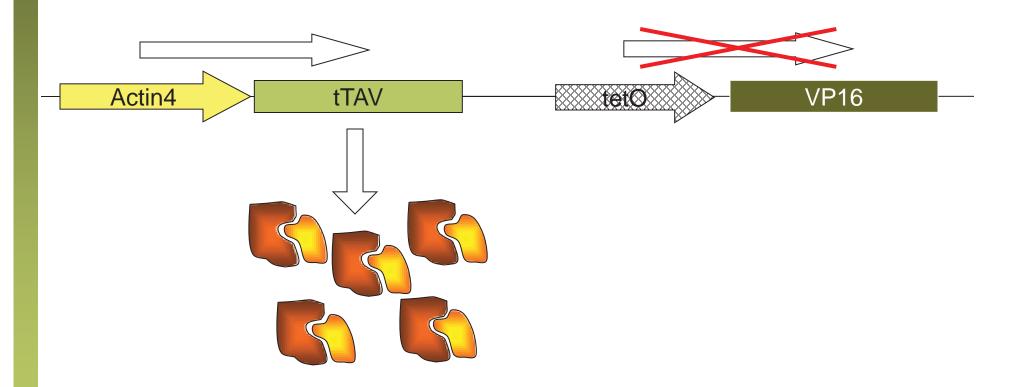
■ Design of the sex-specific construct;



■ This is not a positive feedback system!

Bi-sex RIDL technology

■ Design of the sex-specific construct;



RIDL mosquitoes

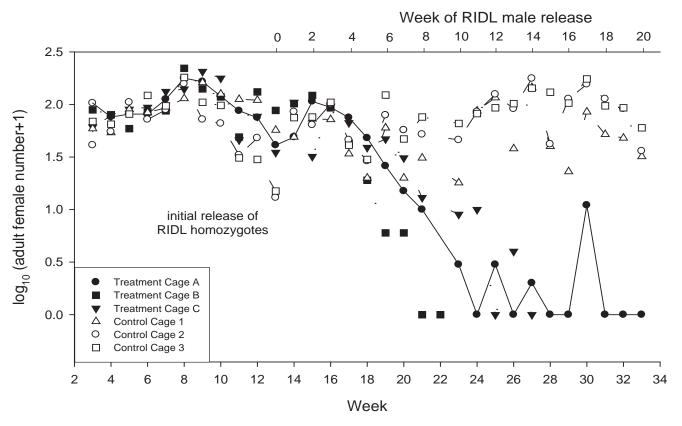




Males Females

Flightless mosquitoes cannot survive in wild (or find hosts). Unable to mate even in laboratory. Males have normal flight ability, as have females given antidote as larvae.

OX3604C Cage suppression trial results



- Colorado State University (Oxitec, Megan Wise de Valdez, Bill Black, Anthony James) experiment in indoor cages
- Control from RIDL was as predicted in simulation model
- Cage populations eradicated in under 20 weeks

Summary:

- RIDL is a safe and effective tool for mosquito control
- Species-specific, environmentally friendly technology
- Under regulatory review for use in the US (Key West trial, commercial licence)
- Applicable to a wide range of species

Acknowledgments



Thank you...